

# The neuroprotective role of L-cysteine towards the effects of short-term exposure to lanthanum on the adult rat brain antioxidant status and the activities of acetylcholinesterase, (Na<sup>+</sup>,K<sup>+</sup>)- and Mg<sup>2+</sup>-ATPase

Charis Liapi · Apostolos Zarros · Stamatios Theocharis ·  
Hussam Al-Humadi · Foteini Anifantaki · Elena Gkrouzman ·  
Zois Mellios · Nikolina Skandali · Stylianos Tsakiris

Received: 17 April 2008 / Accepted: 6 October 2008 / Published online: 21 October 2008  
© Springer Science+Business Media, LLC. 2008

**Abstract** Lanthanum (La) is a rare earth element that is widely used for industrial, medical and agricultural purposes. Its neurotoxic effects are linked to its physical and chemical properties and its interaction with certain trace elements and membrane-bound enzymes. The aim of this study was to investigate the effects of short-term La-administration (as LaCl<sub>3</sub>, 53 mg/kg) on the adult rat whole brain total antioxidant status (TAS) and the activities of acetylcholinesterase (AChE), Na<sup>+</sup>,K<sup>+</sup>-ATPase and Mg<sup>2+</sup>-ATPase, as well as the potential effect of the co-administration of the antioxidant L-cysteine (Cys, 7 mg/kg) on the above parameters. Twenty-eight male Wistar rats were divided into four groups: A (saline-treated control), B (La), C (Cys), and D (La and Cys). All rats were treated once daily with intraperitoneal injections of the tested compounds, for 1-week. Rats

were sacrificed by decapitation and the above mentioned parameters were measured spectrophotometrically. Rats treated with La exhibited a significant reduction in brain TAS (−36%,  $P < 0.001$ , BvsA), that was partially limited by the co-administration of Cys (−13%,  $P < 0.01$ , DvsA), while Cys (group C) had no effect on TAS. The rat brain AChE activity was found significantly increased by both La (+23%,  $P < 0.001$ , BvsA) and Cys (+59%,  $P < 0.001$ , CvsA), while it was adjusted to control levels by the co-administration of La and Cys. The activity of rat brain Na<sup>+</sup>,K<sup>+</sup>-ATPase was significantly decreased by La-administration (−28%,  $P < 0.001$ , BvsA), while Cys supplementation could not reverse this decrease. The activity of Mg<sup>2+</sup>-ATPase exhibited a slight but statistically significant reduction due to La (−8%,  $P < 0.01$ , BvsA), that was further reduced by Cys co-administration (−25%,  $P < 0.001$ , DvsA). The above findings suggest that La short-term in vivo administration causes a statistically significant decrease in the rat brain TAS and an increase in AChE activity. Both effects can be, partially or totally, reversed into control levels by Cys co-administration, which could thus be considered for future applications as a neuroprotective agent against chronic exposure to La. The activities of Na<sup>+</sup>,K<sup>+</sup>- and Mg<sup>2+</sup>-ATPase that were inhibited by La, could not be reversed by Cys co-administration. A role for the already reported concentration-dependent interaction of La with Ca-binding sites (such as Ca<sup>2+</sup>-ATPase) might be considered for certain of the above phenomena.

C. Liapi · A. Zarros · H. Al-Humadi ·  
F. Anifantaki · E. Gkrouzman · N. Skandali  
Department of Pharmacology, Medical School, University  
of Athens, Athens, Greece

A. Zarros · Z. Mellios · S. Tsakiris (✉)  
Department of Physiology, Medical School, University of  
Athens, P.O. Box 65257, 15401 Athens, Greece  
e-mail: stsakir@med.uoa.gr

S. Theocharis  
Department of Forensic Medicine & Toxicology, Medical  
School, University of Athens, Athens, Greece

**Keywords** Lanthanum chloride · Rat brain · Antioxidant status · Acetylcholinesterase ·  $\text{Na}^+, \text{K}^+$ -ATPase ·  $\text{Mg}^{2+}$ -ATPase · L-Cysteine

## Introduction

The rare earth elements (REEs) comprise a group of metallic elements that have similar chemical and physiological properties (Das et al. 1988; Evans 1983). Lanthanum (La) is a REE used for medical (as a treatment of hyperphosphatemia due to chronic renal failure) (Hutchison et al. 2004) and agricultural (as a trace fertilizer) (Guo et al. 1990) purposes. Moreover, La is also released into the aquatic environment through industrial (electronic-manufacturing), mining (of gold and uranium) and energy-producing (through coal-fired power plants) activities (Goetz et al. 1982; Noller 1991, 1994). In fact, it has been suggested that lanthanides (the series of elements relevant to La) are released into the environment in amounts that largely exceed the environmental release of mercury or cadmium by 50 times, of selenium, uranium or tin by 10 times, and of arsenic or lead by 2–3 times (Sabbioni et al. 1982). Thus, the amounts of La that reach the human body, either through the food chain, medical applications or via atmospheric particles, are high (Zhu et al. 1997).

The REEs are not considered as highly toxic for mammals (Haley 1979). However, studies have shown that children living near lanthanides' ore areas tend to have lower IQ levels than those from other regions (Fan et al. 2004; Zhu et al. 1996). A number of experimental studies have focused on the La-induced neurotoxicity, and have revealed some very interesting findings (Basu et al. 1982, 1984; Feng et al. 2006; Gundersen and Miledi 1983). It has been recently suggested that La could possibly impair the learning ability due to a disturbance of the homeostasis between trace elements, enzymes and neurotransmitter systems in the rat brain (Feng et al. 2006). However, this hypothesis has been questioned on the ground of La being an element that can difficultly cross the blood–brain barrier and has questionable concentration-dependent biological effects (Damment et al. 2007).

It should be noted that La has many chemical and physical characteristics in common with calcium (Ca) (Evans 1983), and that its biological actions are

mainly mediated through the displacement or replacement of Ca in certain Ca-binding sites (Das et al. 1988), as well as through its high affinity for the phosphate groups of certain macromolecules (Howells and Coult 1971). Moreover, since REEs are believed to be excluded by the plasma membrane and cannot enter the cytoplasm, the plasma membrane molecules are considered as their primary target (Gao et al. 1998).

The aim of this study was to shed more light on the effects of short-term high-dose La-administration on: (a) the adult rat brain total antioxidant status (TAS) and (b) the activities of acetylcholinesterase (AChE; a crucial membrane-bound enzyme involved in cholinergic neurotransmission) (Kouniniotou-Krontiri and Tsakiris 1989) and two important adenosinetriphosphatases, namely  $\text{Na}^+, \text{K}^+$ -ATPase (an enzyme implicated in neuronal excitability, metabolic energy production, as well as in the uptake and release of catecholamines, serotonin and glutamate) (Bogdanski et al. 1968; Hernandez 1987; Lees et al. 1990; Mata et al. 1980; Sastry and Phillis 1977; Swann 1984) and  $\text{Mg}^{2+}$ -ATPase (an enzyme functioning in order to maintain high brain intracellular  $\text{Mg}^{2+}$ , thus possibly controlling the rate of protein synthesis and cell growth) (Sanui and Rubin 1982). Moreover, since L-cysteine (Cys) is a well known antioxidant and chelating-agent (Carageorgiou et al. 2004; Patrick 2003), it was co-administered with La, in order to evaluate its efficacy on protecting the rat brain against the La-induced (toxic) effects on the above parameters.

## Materials and methods

### Animals

Male adult albino Wistar rats (4 months old, weighting  $279 \pm 25$  g) were used in all experiments. The rats were purchased from the Hellenic Pasteur Institute (Athens, Greece) and were housed four in a cage, at a constant room temperature ( $22 \pm 1^\circ\text{C}$ ) under a 12-h light:12-h dark (light 08:00–20:00 h) cycle. Food and water were provided ad libitum. Animals were cared for in accordance with the principles for the care, use and protection of experimental animals as set by the European Economic Community (EEC) Council Directive 86/609/EEC

(EEC Council 1986) and aligned according to the Recommendation 2007/526/EU.

### Lanthanum and L-cysteine administration

Rats were divided into four groups ( $n = 7$  at each group), as follows: (a) control (saline-treated), (b) La (53 mg/kg of body weight, as  $\text{LaCl}_3 \cdot 7\text{H}_2\text{O}$ , which equals to the  $\frac{1}{2}$  of the  $\text{LD}_{50}$ ), (c) Cys (7 mg/kg of body weight), and (d) La + Cys (on the pre-mentioned doses). All rats received intraperitoneal (ip) daily injections for 7 days. No behavioral or physiological effects were observed over this period of administration.

### Tissue preparation

The animals were sacrificed by decapitation 1 h after the last injection and their whole brains were rapidly removed (and stored at  $-80^\circ\text{C}$  until use). The tissue was homogenized in 10 vol ice-cold ( $0-4^\circ\text{C}$ ) medium containing 50 mM Tris (hydroxymethyl) aminomethane-HCl (Tris-HCl), pH 7.4 and 300 mM sucrose, using an ice-chilled glass homogenizing vessel at 900 rpm (4–5 strokes). Then, the homogenate was centrifuged at  $1,000\times g$  for 10 min to remove nuclei and debris (Tsakiris 2001; Tsakiris et al. 2000). In the resulting supernatant, the protein content was determined according to the method of Lowry et al. (1951) and then the enzyme activities were measured.

### Determination of the brain total antioxidant status

TAS was evaluated in each fresh homogenized rat brain (Carageorgiou et al. 2003, 2005). The total antioxidant capacity was measured spectrophotometrically by a commercially available kit (Randox Laboratories Ltd., Cat No NX2332) as previously described (Tsakiris et al. 2000). Briefly, ABTS (2,2'-Azino-di-(3-ethylbenzthiazoline sulphonate)) was incubated with a peroxidase (metmyoglobin) and  $\text{H}_2\text{O}_2$  in order to produce the radical cation  $\text{ABTS}^{\cdot+}$ . The latter had a relatively stable blue-green color, which was measured at 600 nm. Decreased values of TAS reflect the increase of brain free radical production, whereas increased TAS values show the decrease of free radical production and the protective

antioxidant effect on the brain of the administered substance.

### Determination of brain acetylcholinesterase activity

AChE activity was determined by following the hydrolysis of acetylthiocholine according to the method of Ellman et al. (1961), as previously described by Tsakiris (2001). The incubation mixture (1 ml) contained 50 mM Tris-HCl, pH 8, 240 mM sucrose and 120 mM NaCl. The protein concentration of the incubation mixture was 80–100  $\mu\text{g}/\text{ml}$ . The reaction was initiated after addition of 0.03 ml of 5,5'-dithionitrobenzoic acid (DTNB) and 0.05 ml of acetylthiocholine iodide, which was used as substrate. The final concentration of DTNB and substrate were 0.125 and 0.5 mM, respectively. The reaction followed spectrophotometrically by the increase of absorbance ( $\Delta\text{OD}$ ) at 412 nm.

### Determination of $\text{Na}^+, \text{K}^+$ -ATPase and $\text{Mg}^{2+}$ -ATPase activities

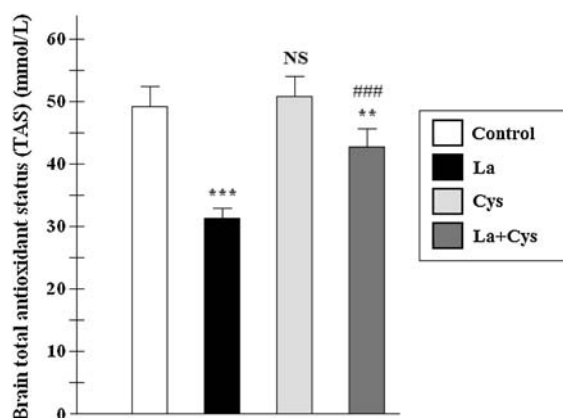
( $\text{Na}^+, \text{K}^+$ )-ATPase activity was calculated from the difference between total ATPase activity ( $\text{Na}^+, \text{K}^+, \text{Mg}^{2+}$ -dependent ATPase) and  $\text{Mg}^{2+}$ -dependent ATPase activity. Total ATPase activity was assayed in an incubation medium consisting of 50 mM Tris-HCl, pH 7.4, 120 mM NaCl, 20 mM KCl, 4 mM  $\text{MgCl}_2$ , 240 mM sucrose, 1 mM ethylenediamine tetraacetic acid  $\text{K}_2$ -salt ( $\text{K}^+$ -EDTA), 3 mM disodium ATP and 80–100  $\mu\text{g}$  protein of the homogenate in a final volume of 1 ml. Ouabain (1 mM) was added in order to determine the activity of  $\text{Mg}^{2+}$ -ATPase. The reaction was started by adding ATP and stopped after an incubation period of 20 min by addition of 2 ml mixture of 1% lubrol and 1% ammonium molybdate in 0.9 M  $\text{H}_2\text{SO}_4$  (Bowler and Tirri 1974; Tsakiris 2001). The yellow color which developed was read at 390 nm.

### Statistical analysis

The data were analyzed by Student's *t*-test followed by Bonferroni post-hoc test when needed. All analyses were performed by SPSS 10.0 statistical package for windows software, while *P* values of  $<0.05$  were considered statistically significant.

## Results

The effects of La and/or Cys on the adult rat brain TAS are presented in Fig. 1. A La-induced statistically significant decrease in TAS was recorded (−36%,  $P < 0.001$ ), that was partially limited by the co-administration of Cys (−13%,  $P < 0.01$ ). The rat brain AChE activity was found significantly increased by both La (+23%,  $P < 0.001$ ) and Cys (+59%,  $P < 0.001$ ), while it was adjusted into control levels by the co-administration of La and Cys (Table 1). The



**Fig. 1** Effects of lanthanum (La) on the brain total antioxidant status (TAS) of adult rats and modulation by L-cysteine (Cys) co-administration. Each value indicates the mean  $\pm$  SD of seven independent experiments (seven rats per group). The average of each experiment arose from three evaluations of the homogenized brain of each animal. NS non statistically significant (as compared to the control group); \*\* $P < 0.01$  (as compared to the control group); \*\*\* $P < 0.001$  (as compared to the control group); ### $P < 0.001$  (as compared to the La-treated group)

**Table 1** Effects of lanthanum (La) on the brain acetylcholinesterase (AChE) activity of adult rats and modulation by L-cysteine (Cys) co-administration

Group ( $n = 7$ )	AChE ( $\Delta\text{OD}/\text{min} \times \text{mg protein}$ )		
	Mean $\pm$ SD	vs. Control	vs. La
Control	0.721 $\pm$ 0.054 <sup>a</sup>	–	–
La	0.890 $\pm$ 0.035 <sup>b</sup>	(+23%)	–
Cys	1.146 $\pm$ 0.031 <sup>c</sup>	(+59%)	–
La + Cys	0.750 $\pm$ 0.041 <sup>d</sup>	(+4%)	(−16%)

Each value indicates the mean  $\pm$  SD of seven independent experiments (seven rats per group). The average of each experiment arose from three evaluations of the homogenized brain of each animal. Statistics: a/b, a/c, b/c, b/d, c/d:  $P < 0.001$ ; a/d:  $P > 0.05$

**Table 2** Effects of lanthanum (La) on the brain  $\text{Na}^+, \text{K}^+$ -ATPase activity of adult rats and modulation by L-cysteine (Cys) co-administration

Group ( $n = 7$ )	$\text{Na}^+, \text{K}^+$ -ATPase ( $\mu\text{mol Pi/h} \times \text{mg protein}$ )		
	Mean $\pm$ SD	vs. Control	vs. La
Control	2.56 $\pm$ 0.28 <sup>a</sup>	–	–
La	1.85 $\pm$ 0.19 <sup>b</sup>	(−28%)	–
Cys	1.47 $\pm$ 0.31 <sup>c</sup>	(−43%)	–
La + Cys	1.86 $\pm$ 0.23 <sup>d</sup>	(−27%)	(+1%)

Each value indicates the mean  $\pm$  SD of seven independent experiments (seven rats per group). The average of each experiment arose from three evaluations of the homogenized brain of each animal. Statistics: a/b, a/c, a/d:  $P < 0.001$ ; b/c, c/d:  $P < 0.05$ ; b/d:  $P > 0.05$

**Table 3** Effects of lanthanum (La) on the brain  $\text{Mg}^{2+}$ -ATPase activity of adult rats and modulation by L-cysteine (Cys) co-administration

Group ( $n = 7$ )	$\text{Mg}^{2+}$ -ATPase ( $\mu\text{mol Pi/h} \times \text{mg protein}$ )		
	Mean $\pm$ SD	vs. Control	vs. La
Control	7.25 $\pm$ 0.31 <sup>a</sup>	–	–
La	6.66 $\pm$ 0.36 <sup>b</sup>	(−8%)	–
Cys	6.13 $\pm$ 0.18 <sup>c</sup>	(−15%)	–
La + Cys	5.46 $\pm$ 0.24 <sup>d</sup>	(−25%)	(−18%)

Each value indicates the mean  $\pm$  SD of seven independent experiments (seven rats per group). The average of each experiment arose from three evaluations of the homogenized brain of each animal. Statistics: a/c, a/d, b/d, c/d:  $P < 0.001$ ; a/b, b/c:  $P < 0.01$

activity of rat brain  $\text{Na}^+, \text{K}^+$ -ATPase was significantly decreased by La-administration (−28%,  $P < 0.001$ ), while Cys co-administration could not reverse this decrease (Table 2). The activity of  $\text{Mg}^{2+}$ -ATPase exhibited a slight but statistically significant reduction in its activity (−8%,  $P < 0.01$ ) due to La, that was further reduced by Cys co-administration (Table 3).

## Discussion

Exposure to La is known to cause a variety of biological effects in a time- and dose-dependent manner (Zheng et al. 2000). Lipid peroxidation of plasma membranes is depressed by low and enhanced by high La concentrations, while the reduction rate of  $\text{Fe}(\text{CN})_6^{3-}$  (a redox system activity marker) is

increased by low and decreased by high La concentrations (Zheng et al. 2000). Our data revealed a statistically significant La-induced reduction of whole rat brain TAS (Fig. 1), following high-dose short-term in vivo administration. The co-administration of Cys was able to partially correct the La-induced TAS-decrease, possibly due to the chelating properties of Cys (assisting to the biological inactivation and/or excretion of La ions) and/or to the possible elimination of the La-induced free radical production. However, Cys (at least under the examined experimental conditions) was not proved efficient enough to neutralize the La-induced oxidative stress.

The co-administration of La and Cys was, on the other hand, efficient enough in order to maintain AChE into the control levels (Table 1) and reverse the La-induced AChE activation. This finding is of importance, since most in vitro studies have reported a La-induced decrease in AChE activity (Ghosh et al. 1991; Gundersen and Miledi 1983; Marquis and Black 1985). Could this finding support the view of Damment et al. (2007) that La can difficultly cross the blood–brain barrier? Could this support the idea that the observed AChE activation is an indirect effect of La? If so, what is the reason of this in vivo AChE activation, and how does Cys manage to reverse it?

Recent studies (Feng et al. 2006; Xiao et al. 2005) have suggested that lanthanides can penetrate the blood–brain barrier when administered in high concentrations, and can accumulate in certain crucial rat brain regions (such as those of the cerebral cortex, the hippocampus and the cerebellum). Moreover, La has been shown to inhibit the Ca-dependent neurotransmitter release (Przywara et al. 1992) and modulate the ion homeostasis within the central nervous system (CNS) (Feng et al. 2006). Since Cys also activates the rat brain AChE, the mechanism by which Cys exerts its neuroprotective effect towards La cannot be directly related to the enzyme, but is possibly taking place through a chelator-metal ion interaction that limits extracellular free  $\text{La}^{3+}$  levels within the CNS and/or restricts the indirect (oxidative or deregulating) effects of La (towards other enzymes and mechanisms of neurotransmission) within the body and/or the CNS. However, this hypothesis requires further investigation, since: (a) it is not clear whether La is directly affecting AChE activity through its physical presence within the CNS (Damment et al. 2007), (b)

little is known about the consequences of the reported ion homeostasis deregulation (Feng et al. 2006), while (c) the mechanisms by which the La-induced cognitive decline is believed to develop do not implicate any change in acetylcholine levels (Feng et al. 2006).

The La-induced decrease of the whole rat brain  $\text{Na}^+, \text{K}^+$ -ATPase activity (Table 2) was expected (Li et al. 1998), while the Cys-induced decrease in  $\text{Na}^+, \text{K}^+$ -ATPase activity has already been reported by Carageorgiou et al. (2004). In contrast to the pre-mentioned parameters (TAS and AChE activity), Cys was not able to reverse this La-induced inhibition of  $\text{Na}^+, \text{K}^+$ -ATPase. This finding suggests that La might interfere with neuronal excitability, metabolic energy production, as well as with the uptake and release of catecholamines, serotonin and glutamate (Bogdanski et al. 1968; Hernandez 1987; Lees et al. 1990; Mata et al. 1980; Sastry and Phillis 1977; Swann 1984), in a way that cannot be reversed by the administration of an antioxidant and chelating-agent, such as Cys. This finding seems to be very important: previous studies have implicated La-administration in glutamatergic, serotonergic and dopaminergic dysfunction (Basu et al. 1984; Feng et al. 2006). Moreover, the ability of La to cause a (slight but significant) decrease of  $\text{Mg}^{2+}$ -ATPase activity, that could also not be reversed by Cys co-administration (Table 3), leads to the conclusion that La also affects the cellular mechanisms functioning in order to maintain high brain intracellular  $\text{Mg}^{2+}$ , and thus possibly controlling the rate of protein synthesis and cell growth (Sanui and Rubin 1982). The observed effects of La on the examined adenotriphosphatases could be the result of  $\text{Ca}^{2+}$ -ATPase inhibition (Basu et al. 1982; Zheng et al. 2000), resulting in Ca-homeostasis deregulation, membrane signaling dysfunction, ATP consumption and/or ion redistribution.

In conclusion, our findings suggest that short-term in vivo La-administration causes a significant decrease in the rat brain TAS and a significant increase in AChE activity. Both effects can be, partially or totally, reversed into control levels by Cys co-administration, which could thus be considered for future applications as a neuroprotective agent against poisoning and/or chronic exposure to La. The activity of  $\text{Na}^+, \text{K}^+$ -ATPase as well as that of  $\text{Mg}^{2+}$ -ATPase were inhibited by La, in a non-reversible by Cys way. These findings suggest that La, at least under the examined experimental conditions, might interfere



with neuronal excitability, metabolic energy production, as well as with the uptake and release of catecholamines, serotonin and glutamate. Some of these phenomena might be the result of oxidative stress, cholinergic dysfunction and/or other intracellular deregulating phenomena associated with its chemical resemblance to other elements such as Ca. However, the matter requires further investigation in order to elucidate the exact mechanisms that lead (or at least have the potential to lead) to the (already reported in the literature) La-induced cognitive impairment.

**Acknowledgments** This study was funded by the University of Athens. Many thanks are expressed to the medical students John Botis, Vasilios Memtsas and Marios Margaritis for their assistance.

## References

- Basu A, Chakrabarty K, Chatterjee GC (1982) Neurotoxicity of lanthanum chloride in newborn chicks. *Toxicol Lett* 14:21–25
- Basu A, Chakrabarty K, Chatterjee GC (1984) The effects of lanthanum chloride administration in newborn chicks on glutamate uptake and release by brain synaptosomes. *Toxicol Lett* 20:303–308
- Bogdanski DF, Tissari A, Brodie BB (1968) Role of sodium, potassium, ouabain and reserpine in uptake, storage and metabolism of biogenic amines in synaptosomes. *Life Sci* 7:419–428
- Bowler K, Tirri R (1974) The temperature characteristics of synaptic membrane ATPases from immature and adult rat brain. *J Neurochem* 23:611–613
- Carageorgiou H, Zarros A, Tsakiris S (2003) Selegiline long-term effects on brain acetylcholinesterase, ( $\text{Na}^+$ ,  $\text{K}^+$ )-ATPase activities, antioxidant status and learning performance of aged rats. *Pharmacol Res* 48:245–251
- Carageorgiou H, Tzotzes V, Pantos C, Mourouzis C, Zarros A, Tsakiris S (2004) *In vivo* and *in vitro* effects of cadmium on adult rat brain total antioxidant status, acetylcholinesterase, ( $\text{Na}^+$ ,  $\text{K}^+$ )-ATPase and  $\text{Mg}^{2+}$ -ATPase activities: protection by L-cysteine. *Basic Clin Pharmacol Toxicol* 94:112–118
- Carageorgiou H, Pantos C, Zarros A, Mourouzis I, Varonos D, Cokkinos D, Tsakiris S (2005) Changes in antioxidant status, protein concentration, acetylcholinesterase, ( $\text{Na}^+$ ,  $\text{K}^+$ )- and  $\text{Mg}^{2+}$ -ATPase activities in the brain of hyper- and hypothyroid adult rats. *Metab Brain Dis* 20:129–139
- Dammit SJP, De Broe ME, D'Haese PC, Bramall N, Cox AG, McLeod CW (2007) Incredulous effects of lanthanum? *Toxicol Lett* 168:186–189
- Das T, Sharma A, Geeta T (1988) Effects of lanthanum in cellular systems. *Biol Trace Elem Res* 18:201–228
- EEC Council (1986) EEC Council Directive 86/609/EEC of 24 November 1986 on the approximation of laws, regulations and administrative provisions of the member states regarding the protection of animals used for experimental and other scientific purposes. *Off J Eur Union* L358:1–28
- Ellman GL, Courtney KD, Andres V Jr, Featherstone RM (1961) A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem Pharmacol* 7:88–95
- Evans CH (1983) Interesting and useful biochemical properties of lanthanides. *TIBS* 8:445–449
- Fan GQ, Yuan ZK, Zheng HL, Liu ZJ (2004) Study on the effects of exposure to rare earth elements and health-responses in children aged 7–10 years. *J Hyg Res* 33:23–28
- Feng L, Xiao H, He X, Li Z, Li F, Liu N, Zhao Y, Huang Y, Zhang Z, Chai Z (2006) Neurotoxicological consequence of long-term exposure to lanthanum. *Toxicol Lett* 165:112–120
- Gao XY, Guo YS, Wu DS (1998) The study on distribution of lanthanum in the rice seedling using the model of cryofixation and cryo-substitution equipment. *Commun Plant Physiol* 34:270–273
- Ghosh N, Chattopadhyay D, Chatterjee GC (1991) Chicken erythrocyte membrane: lipid profile and enzymatic activity under lanthanum chloride and neodymium chloride administration. *Indian J Exp Biol* 29:226–229
- Goetz L, Bignoli G, Sabbioni E (1982) Mobilization of heavy metals from coal-fired power plants: potential impact on ground water in quality of groundwater. *Stud Environ Sci* 17:261–264
- Gundersen CB, Miledi R (1983) Acetylcholinesterase activity of *Xenopus laevis* oocytes. *Neuroscience* 10:1487–1495
- Guo BS, Zhu WM, Xiong BK, Ji YJ, Liu Z, Wu ZM (1990) Rare earths in agriculture. China Agricultural Science and Technology Press, Beijing, p 11
- Haley TJ (1979) Toxicity. In: Schneider KA Jr, Eyring L (eds) *Handbook on the physics and chemistry of rare earths*. North-Holland, Amsterdam, pp 553–585
- Hernandez J (1987) Brain  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity possibly regulated by a specific serotonin receptor. *Brain Res* 408:399–402
- Howells DJ, Coult DB (1971) Ions of the rare earths as possible reactivators of acetylcholinesterase inhibited by some organophosphorus compounds. *Biochim Biophys Acta* 244:427–431
- Hutchison AJ, Speake M, Al-Baaj F (2004) Reducing high phosphate levels in patients with chronic renal failure undergoing dialysis: a 4-week, dose-finding, open-label study with lanthanum carbonate. *Nephrol Dial Transplant* 19:1902–1906
- Kouniniotou-Krontiri P, Tsakiris S (1989) Time dependence of  $\text{Li}^+$  action on acetylcholinesterase activity in correlation with spontaneous quantal release of acetylcholine in rat diaphragm. *Jpn J Physiol* 39:429–440
- Lees GJ, Lehmann A, Sandberg M, Hamberger A (1990) The neurotoxicity of ouabain, a sodium-potassium ATPase inhibitor, in the rat hippocampus. *Neurosci Lett* 120:159–162
- Li XM, Ni JZ, Chen JW, Hwang F (1998) Effects of  $\text{La}^{3+}$  on lipid fluidity and structural transitions in human erythrocyte membranes. *Biochem Mol Biol Int* 45:323–330
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ (1951) Protein measurement with the Folin phenol reagent. *J Biol Chem* 193:265–275
- Marquis JK, Black EE (1985) Activation and inactivation of bovine caudate acetylcholinesterase by trivalent cations. *Biochem Pharmacol* 34:533–538

- Mata M, Fink DJ, Gainer H, Smith CB, Davidsen L, Savaki H, Schwartz WJ, Sokoloff L (1980) Activity-dependent energy metabolism in rat posterior pituitary primarily reflects sodium pump activity. *J Neurochem* 34:213–215
- Noller BN (1991) Non-radiological contaminants from uranium mining and milling at ranger, Jabiru, Northern territory Australia. *Environ Monit Assess* 19:383–400
- Noller BN (1994) The identification of constituents in waste waters from gold mining using ICP-MS. *Int J Surf Min Reclam Environ* 8:95–99
- Patrick L (2003) Toxic metals and antioxidants: part II. The role of antioxidants in arsenic and cadmium toxicity. *Altern Med Rev* 8:106–128
- Przywara DA, Bhavé SV, Bhavé A, Chowdhury PS, Wakade TD, Wakade AR (1992) Activation of  $K^+$  channels by lanthanum contributes to the block of transmitter release in chick and rat sympathetic neurons. *J Membr Biol* 125:155–162
- Sabbioni E, Pietra R, Gaglione P, Vocaturo G, Colombo F, Zanoni M (1982) Long-term occupational risk of rare earth pneumoconiosis. *Sci Total Environ* 26:19–32
- Sanui H, Rubin H (1982) The role of magnesium in cell proliferation and transformation. In: Boynton AL, McKochan WL, Whitfield JP (eds) *Ions cell proliferation and cancer*. Academic Press, New York, pp 517–537
- Sastry BS, Phillis JW (1977) Antagonism of biogenic amine-induced depression of cerebral cortical neurones by  $Na^+$ ,  $K^+$ -ATPase inhibitors. *Can J Physiol Pharmacol* 55:170–179
- Swann AC (1984)  $(Na^+, K^+)$ -adenosine triphosphatase regulation by the sympathetic nervous system: effects of noradrenergic stimulation and lesion in vivo. *J Pharmacol Exp Ther* 228:304–311
- Tsakiris S (2001) Effects of L-phenylalanine on acetylcholinesterase and  $Na^+$ ,  $K^+$ -ATPase activities in adult and aged rat brain. *Mech Ageing Dev* 122:491–501
- Tsakiris S, Angelogianni P, Schulpis KH, Behrakis P (2000) Protective effect of L-cysteine and glutathione on rat brain  $Na^+$ ,  $K^+$ -ATPase inhibition induced by free radicals. *Z Naturforsch [C]* 55:271–277
- Xiao HQ, Li FL, Zhang ZY, Feng LX, Li ZJ, Yang JH, Chai ZF (2005) Distribution of ytterbium-169 in rat brain after intravenous injection. *Toxicol Lett* 155:247–252
- Zheng HL, Zhao ZQ, Zhang CG, Feng JZ, Ke ZL, Su MJ (2000) Changes in lipid peroxidation, the redox system and ATPase activities in plasma membranes of rice seedling roots caused by lanthanum chloride. *Biometals* 13:157–163
- Zhu WF, Xu SQ, Zhang H, Shao PP, Wu DS, Yang WJ, Feng J (1996) Investigation of children intelligence quotient in REE mining area: bio-effect study of REE mining area in South Jiangxi. *Chin Sci Bull* 41:914–916
- Zhu WF, Xu SQ, Shao PP, Zhang H, Feng J, Wu DS, Yang WJ (1997) Investigation on intake allowance of rare earth. A study on bio-effect of rare earth in South Jiangxi. *China Environ Sci* 1:63–66